Usage Policy of the Bioimage Core Facility

The Bioimage Core Facility for Light Microscopy and Image Analysis (BICORE) was founded and is now run by the Department of Biology and the Section of Medicine of the University of Fribourg.

The core facility provides access to shared instrumentation and software for state-of-the-art light microscopy. The core facility staff's mission is to assist you along the path from sample to knowledge with our experience in optimized image acquisition and our expertise in image analysis and data mining.

Who can access the facility?

All research groups from the University of Fribourg and associated Institutes (e.g., Adolphe Merkle Institute) can get access to the facility after registration. With research groups we refer to group leaders or principal investigators (PI) who have her/his own funding (research grants) including the collaborators paid via financial resources of the PI. An official list of research groups is maintained and updated by the administration of each department.

The core facility is also accessible to external users. Using the core facility without an UNIFR informatic account requires additional preparation. However, this procedure is not covered in this document and will be directly communicated to future facility users when needed.

How to get access the facility?

If it is the first contact with the facility, the preferred way is to schedule a meeting with the core facility staff. Future users will be presented with an overview of the services of the facility and get an opportunity to discuss applications and how to best implement them. Invitations from research groups for a meeting with all/several interested lab-members are appreciated.

Once users have decided on which instrument to use for their experiment, the instrument access procedure has to be completed (see get access to an instrument section).

What we do

- Maintain and upgrade a comprehensive set of high-quality optical imaging systems
- Advise users on experimental design and measurement strategies
- Provide training and guidance in the use of the instruments maintained by the core facility
- Maintain and upgrade high-end image analysis software and powerful image analysis workstations
- Operate servers to save and handle imaging data
- Provide all relevant information and a reservation calendar on our website
- Disseminate knowledge on optical imaging through formal teaching, workshops, and hands-on training
- Favour the dissemination of open-source solutions for image acquisition and analysis
- Follow the current development in the field of light microscopy

What we do not do

- Carry out experiments on your behalf
- Perform image analysis on your behalf
- Repair and maintain your 'private' microscope (i.e., instrument not part of the core facility)
Using the facility resources

All facility users have to complete a training session with the facility staff for any given instrument they intend to use. While we encourage colleagues to share their expertise in microscopy with their lab-members, first time users of core facility instruments still have to complete training given by the core facility staff.

Training sessions for all instruments are free of charge. Users can also re-schedule training sessions to refresh their knowledge or to further improve their microscopy skills with a given instrument as needed.

The facility also retains the right to impose subsequent training sessions for microscope users if the facility staff comes to the conclusion that additional instructions are needed or if insufficient proficiency becomes manifest.

Determine the best instrument for the job

When new experiments are planned, we recommend getting in touch with core facility staff members before you submitting the training request. Send us an e-mail \(^1\) with the following information:

- Specimen (cell lines, tissue sections, etc.)
- Staining (fluorophores and their targets, tissue staining, etc.)
- Sample preparation (fixed, live, embedding medium, etc.)
- Carrier (e.g. glass slide, petri-dish, multiwell plate, etc.)
- What is the approximate size of the smallest structure of interest
- What needs to be analysed/quantified

With the above information we can determine which of the instruments is best suited for the task.

Get access to an instrument

1. Submit a training request for a given instrument on the booking system \(^2\) by clicking on the \(\text{!}\) icon of scheduled-resource panel and indicate date/time preferences in the comment field of the request form.

Core facility staff will then schedule the training session (also see IRIS documentation \(^3\) )

2. After completion of the training you will be granted:

   - Access to the booking calendar of the instrument
   - Access to the facility premises (campus card)
   - The right to use the instrument autonomously

Assistance and issue reporting

<table>
<thead>
<tr>
<th>Type</th>
<th>Communication Channel</th>
<th>Examples</th>
</tr>
</thead>
<tbody>
<tr>
<td>Real time assistance</td>
<td>Boris: 8881 Felix: 8545</td>
<td>Software does not start, hardware does not respond, ... something that keeps you from being able to perform your measurements.</td>
</tr>
<tr>
<td>Request to resolve in due time</td>
<td>IRIS (^2)</td>
<td>Refill of consumables needed: used the last cleaning tissue, immersion liquid bottle ran empty, something is missing, c: drive of the computer is almost full ...</td>
</tr>
<tr>
<td>Log book entry</td>
<td>IRIS (^2)</td>
<td>Unexpected, unusual or unexplainable observations: strange noise, little defects on the hardware that do not compromise the entire system ...</td>
</tr>
<tr>
<td>Consulting</td>
<td>e-mail (^1)</td>
<td>Which system is best suited for the type of measurement? How can I optimise my sample preparation? Optimisation of the data acquisition (speed, sampling, etc.). Discussion of measurement strategies. Questions concerning data handling. Help with the data analysis.</td>
</tr>
</tbody>
</table>

To submit an issue use our booking system described IRIS documentation \(^3\). Once logged into IRIS, find the resource you are working with and use the pen-icon \(\text{!}\) in the upper right-hand corner of the instrument panel to open an issue.
Material breaks... and when it does, the core facility staff's concern is not who to blame, but how to fix it swiftly. Reporting issues is the facility users contribution to ensure the smooth operation of the infrastructure. Importantly, failing to report issues in timely manner is treated as negligence.

When reporting an issue please ensure to include the following:

- Short description of the problem
- Minimal, precise steps to reproduce the problem
- Illustrating screenshots/photos/(video) of the problem/error message

For software issues please read the following best practices 4.

**Routine microscope usage**

The microscopes under the care of our core facility are complex and expensive instruments. Please handle them with care. If in doubt, please ask the core facility staff for assistance and advice. Below you find the checklist for each session:

1. Always reserve your time slot (see IRIS documentation 3)
2. Visual check of the system (no obvious defect, make sure to work with clean objectives for best results)
3. Start the system (see manual pages on bioimage wiki 5)
4. Do you measurements
   (soak up the immersion liquid on the objective with a Kimwipe between slide changes)
5. Save the data on a secure location (see section on data management)
6. Shutdown the system (see manual pages on bioimage wiki 5)
7. Clean the objectives with lens cleaning paper and alcohol
8. Clean the work surfaces with alcohol and (normal) cleaning tissue
9. Order for refills if necessary (cleaning utensils, immersion liquids)
   report irregularities (see section on assistance and issue reporting)
Access to facility premises

Access to the core facility premises is controlled by locks with card readers. The facility together with SELOG established a procedure to program the campus card depending on the instruments that need to be accessed. Once the UNIFR account has granted the access, the campus card needs to be updated at one of the campus card machines.

University staff

After completed training, the core facility staff will contact SELOG directly to program the additional door access for the new facility users.

Bachelor and master students

Students without a work contract need to fill out a form and have it signed by their group leader (PI). Once it is filled out it needs to be sent directly to the UNIFR administration. After completion of the training and upon request, the form (with the door numbers pre-filled) can be obtained from the facility staff.

Mailing lists

The core facility uses several mailing lists for different information channels:

- The list bioimage@unifr.ch is used for important announcements to facility users (demos, courses, new equipment, etc).
- The list bioimage-admin@unifr.ch is to address the entire core facility staff.
- The other lists starting bioimage-* can and should be used by microscopy users and allows all members to inform each other.

Facility users are encouraged consult the university’s mailing list server and subscribe to the lists of interest.

Booking rules

Booking rules were implemented to give facility users equal chance to access the instruments and ensure concise booking slots.

1. Every user books his/her own slots.
2. During office hours – Monday to Friday from 9:00 - 17:00 – the following applies:
   - An oh-slot (office hour slot) has, for counting purposes, a maximal duration of 4 hours. E.g. single oh-slots are 9:00 - 13:00, 13:00 - 17:00 but also 7:00 - 12:00. A reservation from 8:00 - 13:30 would count as two oh-slots.
   - At any given time you are allowed maximally three upcoming oh-slots.
   - If possible avoid several slots during office hours on one single day.
3. Outside of the office hours there is no limitation for the duration of the reservation slots.
4. Cancellation must be done before your booked slot has started. Once the slot started booking cancellation becomes impossible and the normal usage fees apply.
5. If you finish your session earlier than scheduled, please inform the next user or all the instrument users via mailing lists (see mailing lists section).
**Usage fees**

To cover the running costs of the light microscopy facility the use of its instruments is subject to the payment of hourly fees. Fees are charged if the instrument is not attended and the booking has not been cancelled at least the day before the reservation date.

The billing is calculated for all members of each research group (PI) and invoices are sent at the end of each trimester to the head of the laboratory.

We distinguish three user types (UT):

- **UT-I**: Platform internal -> Department of Biology and Section of Medicine
- **UT-U**: University internal -> All the staff of the University of Fribourg and associated Institutes
- **UT-E**: External -> All persons not associated with the University of Fribourg

The fees listed in the following table, comprise direct costs (consumables, salaries, maintenance fees) and indirect costs (rent, admin., etc.). For detailed explanations about the cost calculations and what budget you can use to pay the fees (SNF, EU, etc) please contact the core facility.

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<tbody>
<tr>
<td><strong>IC-1.1 Confocal</strong></td>
<td>Leica TCS SP5 (Med) Leica TCS SP5 (Bio) Leica TCS SPE-II</td>
<td>15.00</td>
<td>250</td>
<td>30.00</td>
<td>60.00</td>
</tr>
<tr>
<td><strong>IC-1.2 Cutting-edge confocal</strong></td>
<td>Leica STELLARIS 8 FALCON</td>
<td>20.00</td>
<td>250</td>
<td>35.00</td>
<td>65.00</td>
</tr>
<tr>
<td><strong>IC-2 (Live-Cell)</strong></td>
<td>Visitron CSU-W1 (Spinning Disk Confocal) GE DeltaVision Elite Leica DMI600B (Bio)</td>
<td>7.50</td>
<td>250</td>
<td>25.00</td>
<td>45.00</td>
</tr>
<tr>
<td><strong>IC-3.1 Slide-scanner fluo</strong></td>
<td>Leica DM6B Navigator, Hamamatsu NanoZommer S60</td>
<td>5.00</td>
<td>250</td>
<td>10.00</td>
<td>15.00</td>
</tr>
<tr>
<td><strong>IC-3.2 Slide-Scanner</strong></td>
<td>Hamamatsu NanoZoomer 2.0 HT</td>
<td>0.00</td>
<td>-</td>
<td>15.00</td>
<td>25.00</td>
</tr>
<tr>
<td><strong>Miscellaneous</strong></td>
<td>Marcoscopes, Image Processing Workstations, etc.</td>
<td>0.00</td>
<td>-</td>
<td>0.00</td>
<td>NA</td>
</tr>
<tr>
<td><strong>Data storage</strong></td>
<td>OMERO 8 and HRM 9 use storage on the bigdata server. The fees are according to the rates of the DIT 10 (charged per TB and year)</td>
<td>170</td>
<td>-</td>
<td>170</td>
<td>NA</td>
</tr>
</tbody>
</table>

The **yearly ceiling** defines the maximum amount of cumulative fees per year a single research group is charged to use a given instrument category. For example research group X that uses confocal microscopes 100 hrs/year is charged 1500 CHF. Research group Z that uses confocal microscopes 300 hrs/year is charged 3750 CHF.

NA = Not available
Laboratory safety

The future facility user acknowledges to have studied the information and policies of the SCIMED Faculty.

Laser safety

Certain of our systems use lasers for illumination, photo-bleaching or photo-ablation. Users need to make themselves aware of the safety precautions provided in the manuals of these systems. While the safety mechanisms of the microscopes give a good protection, users need to be careful of mirror-like reflections that their samples might cause.

Data management and data safety

Control computers of any of our instruments are not intended for data storage. Use file servers or external hard drives to store your data (also see our wiki for more detailed information). A full hard drive compromises the effective operation of the instrument. To keep the systems running data saved on the PC hard drive will be deleted on a regular basis and without prior notice. As a consequence, to ensure the data safety, once you leave the microscope the data should be transferred from the microscope's computer to an appropriate storage medium.

Data security

Sensitive data (e.g., samples from human patients) need to be treated according to data protection laws. Before bringing such samples for measurements with core facility instruments, users have to assert that those samples are anonymised.

 Certain computers have large hard drives, accessible by all users of that machine that serves as a buffer space between acquisition and transfer to a permanent data storage. To avoid accidental sharing of sensitive datasets, even anonymised, those data need to be removed from the microscope computers and image processing workstations immediately after their acquisition/processing.

For projects handling such data, make sure that the risk analysis provided by the university's security officer (it-security@unifr.ch) and the security measures are known and implemented.

Accreditation

Core facilities provide the wider research community with state-of-the-art instrumentation and expertise, which not only adds to, but also improves research outputs and funding bids. It is important to recognise the contributions of facility staff to the advancement of scientific research, both formally and informally, in all instances of technical assistance. The type of recognition that is most appropriate will vary dependent upon the contribution provided.

1. All publications resulting from the use of instruments within the facility should, at the very least, acknowledge the as a whole, e.g. *the authors gratefully acknowledge the Bioimage Core Facility of the University of Fribourg for their support & assistance in this work*

2. Where users have had significant help from a particular member of core facility staff or if a core facility staff member has generated additional data for the user, this staff member should be acknowledged by name, alongside the center.

3. If scientists from the core facility contributed more than just routine techniques, they should be co-authors on publications that use such data. For example:
   - Development or adaptation of protocols to suit samples or materials, (re)designing experiments, extensive data analysis and interpretation that are not routine and constitute contributions meriting co-authorship.
   - Implementation of the image processing pipeline, contributing code, and custom software tools (scripts, Jupyter notebooks, imageJ plugins, etc.)

Please send a reprint of the paper, or an e-mail including the reference information for the publication, to core the facility. We also politely request that you remember to acknowledge the use of the core facility in your presentations and cite core facilities in grants, as appropriate.
Duties of the core facility user:

- Follow the directives described in this document
- Provide a cost center who’s owner (PI) agrees to cover the charges of the Bioimage Core Facility
- Be informed (documentation, announcements, etc. on the website)
- Communicate with the core facility staff (see issue tracker, mailing lists, booking rules and accreditation)
- Email a signed copy of this document to the Bioimage Core Facility

I took note of the service scope provided by the Bioimage Core Facility and I agree to the terms of usage described in this document. I understand that bad conduct and breaking these terms can result in the withdrawal of my access to this core facility.

First name: Last name: Digital signature:

1. bioimage-admin@unifr.ch
3. https://www.unifr.ch/scimed/facilities/resources/iris
4. https://imagej.net/Bug_reporting_best_practices (sections 1-4)
5. https://bioimage-wiki.unifr.ch (intranet only)
6. https://www.unifr.ch/campuscard/de/geraete/standorte.html
7. https://lists.unifr.ch/sympa/lists#b
8. https://omero-web.unifr.ch
9. https://bioimage-hrm.unifr.ch
10. http://www.unifr.ch/dit
12. Adapted from the “Publication Core Facility Policy” by Natasha Stepe of the Plymouth Electron Microscopy Centre
13. https://www.unifr.ch/scimed/facilities/facilities/bioimage